FLOW CYTOMETRY CENTER

AUBURN UNIVERSITY COLLEGE OF VETERINARY MEDICINE
FLOW CYTOMETRY & CELL SORTING FACILITY
Room 250A, Department of Pathobiology, Greene Hall
(334) 844-2711

SUBMISSION FORM

***Please fill out as completely as possible.**

Submitted by: _____ Email:_____

Dept: _____ Phone#:_____

What exactly do you want to measure?

- o Analysis ☐ - _____
- o Cell Sorting ☐ - _____

What kind of information are you looking for? (i.e. Surface markers, DNA content, 1 or more colors, 4 way sorting, sorting to 96 well culture plates)

Type of sample(s) submitted: (i.e. cell type/source/species)

Any special preparation (e.g. enrichment, deletion of certain populations, transfection, etc.)

Dye(s) used (i.e. PE, FITC, Alexa):

Wavelength(s)/colors you are interested in: Emission:

Excitation: 488 - ☐ 635 - ☐

Are your cells less than 70 microns?

How many samples are you submitting?

Is your sample fixed (rendered harmless)? _____ Biohazard level: _____

Is your sample TOXIC? (i.e. contains formaldehyde): _____

Have you included all the necessary CONTROLS?

- o Unlabelled cells
- o Isotype controls for each antibody
- o Single label controls for multi-labeled specimens

Please list your controls:

Approximate volume/concentration?

What are your cells suspended in? (i.e. Hanks with 2% FBS, PBS, etc.)

The cells were filtered to:

- o 20 um - ☐
- o 50 um - ☐
- o 70 um - ☐
- o Other - _____

Special Requirements? i.e. Sterile, Purity, Recovery: _____

GUIDELINES

1. Samples must be in 12x75 mm polypropylene test tubes (Falcon #2063) due to the vacuum system in the sampler.

2. ALL samples must be filtered. See website for suggestions.

3. Minimum sample volumes are 0.5 ml.

4. If you have samples stained with more than one fluorochrome per tube, please bring in a sample stained with each fluorochrome individually. This is for compensation purposes.

5. If you are sorting cells should be concentrated – 5 to 10 million cells per ml and FILTERED. Lower concentrations can be sorted but will take additional time. Collection tubes for sorts should be 12x75 Falcon polypropylene tubes with interiors coated with 0.5 ml of 100% FBS. The FBS reduces cell adherence to the walls of the tubes when sorted and keeps the cells HAPPY when they are diluted with sheath fluid. If you are having cells sorted, be prepared to collect them promptly. Sorting dilutes cells greatly so sufficient numbers of tubes must be prepared to collect the sorted cells.

6. Yields of sorted cells are proportional to the % of the population you are sorting for. The time for sorts is proportional to the purity/enrichment desired for that population. If you count your cells immediately prior to sorting the efficiency of yield can be estimated and may also help trace problems associated with low yield.

7. Tubes with visible cell clumping/debris will not be run.

8. The MoFlow is equipped with a temperature controlled sample chamber and collection holder to keep both the pre- and post-sorted cells at a constant temperature. If you would like to use this feature please let the operator know in advance of the sort to equilibrate the equipment to the desired temperature.

9. Tube changes during the sort (both sample and collection) provide opportunities for contamination. Please use antibiotics subsequent to the sort if you are culturing your cells.

Experimental Samples Submitted

Experiment Name:_____ Date:_____
Person Submitting:_____Phone:_____

Tube #	Sample information/conditions	Fluorochrome/probe	Control
1	Unstained cells	NONE	AutoFlourescence control
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